

## **EFFECT OF FOLIAR APPLICATION OF SOME GROWTH PROMOTRES ON GROWTH, FRUITING AND FRUIT QUILITY OF “SULTANI” FIG TREES**

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### **ABSTRACT**

**This investigation was carried out during 2009 and 2010 growing seasons on ten years old Sultani fig trees, grown in a calcareous soil in a private farm in Burg El-Arab region, about 70 kilometers west of Alexandria in order to study the effect of foliar spray with different concentrations of some growth promoters i.e. EM (effective microorganisms) at 10%, 20% and 30% or GA<sub>3</sub> (growth regulator) at 10ppm,20ppm and 30ppm on vegetative growth, fruit set, yield, yield increment, fruit quality and leaf mineral content during both seasons of the study. Results showed that vegetative growth ,fruit set, yield as weight or number of fruits / tree as well as fruit quality (average fruit weight, diameter, length, TSS and total sugar) and leaf N, P and Fe content were generally, improved by increasing the concentration of both treatments as compared with control in both seasons while, slight effect on acidity was noticed. The studied treatments did not significantly affect leaf K, Mn, Zn and fruit vitamin C. in both seasons. The obtained results indicated that 30% of EM was the best concentration to increase all vegetative growth parameters, leaf chlorophyll, N, P and Fe as well as fruit set %, yield and its increment. Similar trend was attained by spraying trees with GA<sub>3</sub> at highest concentration (30ppm) on some fruit quality parameters (fruit**

**weight, length, diameter and total sugar) in both seasons and for TSS in the 1<sup>st</sup> one. It is worthy to note that, spraying trees with highest concentration of EM (30%) had a similar effect to spraying trees with GA<sub>3</sub> at highest concentration (30 ppm). Hence, it could be concluded that spraying fig trees with EM at higher rate (30%) is a promising treatment under the same condition of our study as it is safe, natural, less polluting and recommended to produce highest yield with best fruit quality.**

## INTRODUCTION

In conventional agriculture, chemical fertilizers, pesticides and plant growth regulators are usually applied to increase the yield and quality of crops. However, frequent and excessive use of these chemicals has often resulted in adverse environmental effects, disturbing the ecological balance and making plants even more susceptible to pests and diseases (Bhanti and Taneja, 2007). There is a growing concern that food produced under such farm management may not be safe or of good quality. Public awareness to these problems has shifted the approach towards some alternative measures (Shaxson, 2006). In recent past, bio-product and microbial inoculants have been introduced to modern agriculture as a natural substances to produce food with good quality, and safety with minimize the bad effects of the former (Kannaiyan, 2002). A microbial inoculant containing many kinds of naturally occurring beneficial microorganisms (photosynthetic bacteria, lactic acid, actinomycetes, yeast and fungi) called Effective Microorganisms (EM) according to Higa, 1995 , produced by EMRO corporation, Okinawa Japan and locally marketed by the Ministry of Agric. and Land Reclamation, Egypt.

EM (Effective Microorganisms) has been used widely as inoculants to change the microbial diversity and interaction in soils and plants (Xu, 2000). In turn, EM has been shown to improve soil health, and the growth, yield and quality of crops over a wide range of agro-

ecological conditions (Higa and Parr, 1994; Iwaishi, 2000 and Yamada et al., 2000). When EM cultures are applied to soil they stimulate the decomposition of organic wastes and thereby residues releasing inorganic nutrients for plant uptake.

Foliar application of EM appears to suppress the occurrence of plant diseases and facilitates the uptake of simple organic molecules that can increase plant growth and yield in relatively short time (Wididana and Higa, 1998). Many countries around the world such as Indonesia have come to consider EM in the same way they would apply any foliar fertilizer, especially micronutrients which responded to foliar application of micronutrients much greater than soil application, and believe that foliar application is more efficient (Javaid, 2010). Furthermore, in China, using EM as a foliar application improved the quality and enhanced yield of tea, cabbage and sugar corn (Xiaohou *et al.*, 2001). Moreover, foliar application of EM results in a large number of beneficial microorganisms at the leaf surface, or phyllosphere. It is believed that certain microorganisms in the EM culture including photosynthetic bacteria and N-fixing bacteria, can enhance the plant's photosynthetic rate and efficiency and its N-fixing capacity as well (Pati and Chandra, 1981). Through foliar application, microorganisms in EM appear to suppress the development of harmful plant pathogens at the surface, thereby providing a measure of plant protection through biocontrol. Another example of the beneficial effect of phyllosphere microorganisms was reported by Atlas and Bartha (1981). They found that pigmented yeast and bacteria that colonized on the leaf surfaces could afford some protection to the plant from excessive exposure to direct sunlight. Chamberlain and Daly (2005) reported that the metabolites developed by micro-organisms are directly absorbed into plant surface. In addition photosynthetic bacteria play the leading role in the activity of EM. They synthesize useful substances and increase the number of other bacteria and act as nitrogen binders.

In Egypt, growers sprayed some growth hormones to induce growth and increase fruit set and yield. Gibberellin ( $GA_3$  i.e. Berlex) is the most isomer widely used. They regulate growth and influence various developmental processes, including stem elongation, germination,

flowering, sex expression, enzyme induction and leaf and fruit senescence (Donald et al., 2001). The effect of GA<sub>3</sub> has at least three important actions, intensifies an organ ability to function as a nutrient sink, ability to increase the synthesis of IAA in plant tissues and involves synthesis acceleration of hydrolytic enzymes as amylase and other hydrolytic enzymes in aleurone cells (Addicott and Addicott, 1982). As the effect of growth regulator spray, many investigations on some fruits were accepted from Abd-Ella and El-sisi (2006) on figs and Malaka (2008) on pears.

The objective of the present investigation was to study the effect of foliar applications of EM as a natural stimulator and GA<sub>3</sub> as a growth regulator on fruit set, yield, fruit quality and leaf mineral content of Sultani fig trees.

### **MATERIALS AND METHODS**

This investigation was carried out through two successive seasons 2009 and 2010 on about ten years old Sultani fig trees, grown in a calcareous soil in Burg El Arab region ,about 70 kilometers west of Alex. and spaced at 5x5 apart meters under drip irrigation. Some physical and chemical properties of such soil are listed in Table (1).

The trees in this orchard were annually fertilized with 15 m<sup>3</sup>/feddan of organic manure in December of each year and 1.5 Kg of calcium super phosphate (15.5% P<sub>2</sub>O<sub>5</sub>).Moreover, 3.0 Kg ammonium sulphate (20.5% N) and 1.5 Kg of potassium sulphate (48% K<sub>2</sub>O) per tree were added as soil application in three equal doses at March, April and June. The selected trees were nearly similar in vigor and free from visible pathogens. The trees were sprayed with different treatments in the two seasons as follow:

T<sub>1</sub>: Foliar spray with water only (control).

T<sub>2</sub>: Foliar spray with EM at 10%.

T<sub>3</sub>: Foliar spray with EM at 20%.

T<sub>4</sub>: Foliar spray with EM at 30%.

T<sub>5</sub>: Foliar spray with GA<sub>3</sub> at 10ppm.

T<sub>6</sub>: Foliar spray with GA<sub>3</sub> at 20ppm.

T<sub>7</sub>: Foliar sprat with GA<sub>3</sub> at 30ppm.



Trees were sprayed with the above treatments, three times at full bloom (March), after fruit set (fruit diameter about 2mm) at May and after one month of fruit set (July). Foliar sprays were applied using a hand pressure sprayer. Triton B emulsifier at rate of 0.2% was as a surfactant. Each tree received 5 liters spraying solution. Microbiological and chemical analysis of Effective Microorganisms (EM) is listed in Table (2).

**Table(2) Microbiological and chemical analysis of EM:**

Microbiological Analysis	Population (CFU ml <sup>-1</sup> )	Chemical Analysis	Chemical Content
Total plate count	1.2x10 <sup>9</sup>	N	0.47%
E Coli	0	P	<0.1ppm
Other coliforms	0	K	0.22ppm
Salmonella sp.	0	B	<0.57ppm
Staphylococcus sp.	0	S	<0.1ppm
Clostridium spp.	0	Fe	51ppm
Enterobacter	0	Cu	1ppm
N-fixing bacteria		Mo	<0.03ppm
a. Azospirillum sp.	0	Co	<0.2ppm
b. Azotobacter sp.	0		
c. Rhizobium sp.	0		
P-Solubilizing bacteria	2.0x10 <sup>3</sup>		
Lactobacillus spp.	2.0x10 <sup>6</sup>		
Yeast	6.2x10 <sup>5</sup>		
Fungi	70		
Cellulolytic bacteria	4.3x10 <sup>3</sup>		
Streptomyces sp.	8.6x10 <sup>3</sup>		

CFU = Colony Forming Unit

Treatments were arranged in a Randomized Complete Block Design with three replications for each treatment using three trees as a single replicate (7 treatments x 3 replicates x 3 trees= 63 trees).The following parameters were determined in the two successive seasons:

**1. Vegetative growth:**

At the beginning of both growing seasons, on mid March, eight main branches well distributed around periphery of each replicate tree were randomly selected and tabulated (2 branches toward each direction). Number of the current shoot per selected branches was counted, their lengths and diameters were measured (cm) when the growth was ceased (October 30). Five leaves were collected randomly from the first fully mature leaves from the tip of the previously tagged branches and their areas (cm<sup>2</sup>) were measured.

**2. Leaf chemical components:**

-Leaf chlorophyll reading was recorded using Minolta Chlorophyll Meter Spad-502 (Minolta Camera Co., LTD Japan) in the field at the end of July. The average of ten readings was taken on the middle of leaves from all the tree canopy.

-Leaf mineral content was determined at the end of July of both seasons. Sample of ten leaves was randomly selected from the middle part of non fruiting shoots of each replicate tree in both seasons. Leaf samples were washed with tap water, rinsed with distilled water and oven dried at 70 °C to a constant weight and then ground. The ground samples were digested with sulphuric acid and hydrogen peroxide according to Evenhuis and DeWaard (1980). Suitable aliquots were taken for determination of N, P, K. Nitrogen and phosphorous were determined colorimetrically according to Evenhuis (1976) and Murphy and Riley (1962), respectively. Potassium content was determined against a standard by a Flame photometer according to Chapman and Pratt (1961) while, iron, zinc and manganese by a Perkin Elmer Atomic Absorption Spectrophotometer. The concentrations of N, P and K were expressed as percent, while those of Fe, Zn and Mn were expressed as parts per million, on dry weight basis.

Data were statistically analyzed according to Snedecor and Cochran (1990) and L.S.D test at 0.05 levels was used for comparison between means of various treatments.

### **3. Fruit set percentage and yield:**

The total numbers of buds were recorded and then numbers of syconia on the selected shoots were counted to calculate the percentage of fruit set. Total yield (Kg) of each replicate tree was calculated using average fruit weight (gm) and the total number of fruits per tree. On mid July in both seasons, number of fruits per each experimental tree was counted. Increment percentage compared with the control was also calculated by using the following equation:

$$\frac{\text{Yield (Kg)/ treatment t- yield (Kg)/ control}}{\text{Yield (Kg)/ control}} \times 100$$

### **4. Fruit quality:**

At harvesting time, 1<sup>st</sup> August, of both seasons, ten fruits were taken at random from each replicate tree to determine fruit quality (average fruit weight (gm), diameter (cm) and length (cm)). In juice of each fruit sample, total soluble solids (TSS) percentage was determined by a hand refractometer and the percentage of acidity was measured according to A.O.A.C. (1995). Vitamin C was determined by titration with dichlorophenol endophenol blue dye and expressed as mg vitamin C / 100 ml juice. Total sugars in fruit pulp tissues were determined by phenol sulfuric method according to (Dubois *et al.*, 1956)

## **RESULTS AND DISCUSSION**

### **1-Vegetative growth characters:**

The data representing the effect of foliar application of different concentrations with EM (Effective Microorganisms) and GA<sub>3</sub> (Gibberellic Acid) promoters on vegetative growth parameters were listed in Table (3). In general, the results indicated that, both types of growth promoters either EM or GA<sub>3</sub> was predominant to the control, of all the



evaluated vegetative growth parameters, i.e., number of new shoots per main branch, new shoot length, shoot diameter and leaf area in both seasons. In other words, a gradual increase in the studied growth parameters of "Sultani" fig tree was quite obvious with increasing the concentrations of both EM and GA<sub>3</sub>.

It was clear that foliar application of EM at the highest concentration (30%) was superior and associated with highest value of all previously mentioned characters. The enhancement of plant growth by the EM biostimulant application may be attributed to the profound effect of natural plant growth regulator substances produced by the effective microorganisms (bacteria, fungi, and yeast) as reported by Joo, *et al.*, (1999). Natural hormones like cytokinin which enhance cell division and cell enlargement so far increasing the extension of leaf surface area as well as enhancing the accumulation of soluble metabolites (Ferguson *et al.*, 1987). Moreover, Higa and Wididana (1999) reported that when EM applied to soil or plant leaf surface, the population of photosynthetic bacteria and nitrogen fixing bacteria dramatically increased. This phenomenon was associated with the growth of more vigorous plants and enhanced the plant's photosynthetic rate, efficiency and its nitrogen fixing capacity. In this regard Ruinen (1970) was among the first to investigate the occurrence of nitrogen fixing bacteria on leaf surfaces. The present results were in harmony with those reported by Sangakkara and Marambe (1998) on tomato and french beans and Desoky *et al.*, (2001) on Papaya, who found that trees became more vigorous when subjected to different rates and application methods of foliar sprays with EM. Similarly, Piotr and Zofia (2008) found that apple rootstocks gave significantly greater mass and length of lateral shoots as well as enhanced their diameter and number when sprayed with EM in comparison with water only. While, Javaid and Shah (2010) found that EM foliar application didn't affect various growth parameters of wheat plants.

Data also indicated that foliar application of GA<sub>3</sub> at higher concentration had similar effect on improving some growth parameters (leaf area and shoot diameter). However, the differences between higher

concentrations of both promoters were not big enough to be significant. It is well known that foliar application of GA<sub>3</sub> has the ability to stimulate plant growth and development in a variety of test systems. GA<sub>3</sub> increased plant size as a result of increased photosynthetic rates or due to more efficient utilization of photosynthetic products (Erkan and Bangerth, 1980). El-Sabagh and Mostafa (2003) revealed that an increase in total area of leaves in response to foliar spray of GA<sub>3</sub>. The results were in agreement with some investigators who worked on the effect of GA<sub>3</sub> on apple trees; Grochowska *et al.*, (1995), Callejas-R *et al.*, (1998) and El-Sabagh and Mostafa (2003).

### **2- Leaf chemical components:**

The results presented in Table (4), clearly, indicated that leaf chlorophyll, N, P and Fe were increased by foliar application of EM (commercial bio stimulant) or GA<sub>3</sub> compared to the control in both seasons, while, leaf K, Mn and Zn were not significantly affected. In general, it was noticed that the highest levels of leaf chlorophyll, N, P and Fe were obtained from trees sprayed with highest concentration of EM (30%) in both seasons. The promoting effect of natural bio stimulant on the nutritional status of the leaves could be related to the role of the effective microorganisms in improving the availability of nutrients. Similar findings were recorded by Sangakkara and Nissanka (1998) who found that foliar application of EM significantly increased leaf chlorophyll content and enhanced yield of beans due to greater rates of photosynthesis.

### **3-Fruit set percentage and yield:**

The results of foliar treatments of EM (natural stimulant) and GA<sub>3</sub> (growth promoter) on fruit set and yield of Sultani fig trees were shown in Table (5). It was clear that fruit set and yield as number or weight (Kg/tree) of fruits were significantly increased with increasing the concentration of both EM and GA<sub>3</sub>. Data in the same table indicated that all treatments had a highly significant effect on percentage of yield increment as compared to untreated trees (control) in both seasons. However, spraying trees with EM at high concentration (30% ) gave





significantly the highest fruit set. Similarly, spraying trees with high concentration of GA<sub>3</sub> (30 ppm) gave almost the same effect as high concentration of EM on number of fruits / tree in the 2<sup>nd</sup> season and on other fruiting aspects in the 1<sup>st</sup> one. These results reflect similar trends to those of plant growth and mineral content of leaves as previously mentioned. Therefore, increasing Sultani fig yield might be attributed to the increments in the amounts of metabolites synthesized by the plant which, in turn accelerated plant growth and resulted in improving total yield. These results can be explained as the EM biostimulant contains more than 60 strains of microorganisms as bacteria, yeast, actinomycetes and various fungi. The high contents of minerals and vitamins as well as the cytokinin contents in yeast might play a role in the orientation and translocation of metabolites from leaves into the productive organs (Nagodawithana,1991). Similar results were recorded by Wididiana and Higa (1998), Sangakkara and Marambe (1998) and Xiaohou *et al.*, (2001) on various vegetables crops , Yousaf *et al.*, (2000) on groundnut and Desoky *et al.*, (2001) on papaya , they clearly indicated that foliar solution of EM at certain concentrations and time intervals caused significant increase in yield . Additionally, EM can be used as a regulation substance to improve metabolism of crop plants for yield promotion and quality improvement. Also, these results may be due to spraying trees with plant hormones (GA<sub>3</sub> at 30ppm) which may stimulate shoot growth and there is a strong competition and relationship between the developing fruitlets and rapidly growing shoot tips (El-Sabagh and Mostafa,2003). GA<sub>3</sub> plays a major role in enlarging fruit size (Wiltank and Krezdorn, 1969). These results are in harmony with those obtained by Abd-Ella and El-Sisi (2006) on fig and Malaka (2008) on pear.

#### **4-Fruit quality:**

The results in Table(6) showed gradual and significant increase of fruit physical and chemical properties i.e. fruit weight, diameter, length ,TSS and total sugar % with increasing the concentration of both EM and GA<sub>3</sub> treatments as compared with control. Acidity, however showed an opposite trend, whereas, V.C. was not significantly affected.



It is noteworthy to mention that the effect of spraying trees with GA<sub>3</sub> at higher concentration (30ppm) was more pronounced than other treatments followed by 30 % of EM. However, the trees sprayed with tap water were the lowest. In addition, the response of fig trees to high concentration of GA<sub>3</sub> was almost as like as that of high concentration of EM on fruit juice V.C., besides decreasing acidity. Significant differences were found between the effects of the highest two concentrations of GA<sub>3</sub> and EM on fruit weight in the 2<sup>nd</sup> season and fruit length and diameter in both seasons of study. In the meantime, the differences between the two growth promoters at higher concentration were too few to be significant in TSS and total sugar during both seasons. Apparently, the gibberellins affect cell elongation and cell division, therefore GA<sub>3</sub> plays a major role in enlarging fruit size. The results were in line with those obtained by El-Sabagh and Ahmed (2004) on apple, Abd-Ella and El-Sisi (2006) on fig and Malaka (2008) on pear. The above mentioned results of EM on fruit quality was due to its content of bacteria and yeast which, via its cytokinin content might play a role in the synthesis of protein and nucleic acids and minimized their degradation (Legocka,1987). Likewise, Wood *et al.*, (1997) reported that foliar spray with EM produced plant hormones, beneficial bioactive substances, and antioxidants which solubilize nutrients. Similarly, Shou-Song *et al.*, (2002) suggested that EM can be used as a regulation substance to improve metabolism of crop plants for yield promotion and quality improvement. These results are in line with those obtained by Higa and Wididana (1999) on green pepper and turnip and Desoky *et al.*, (2001) on papaya. They found that foliar application with EM improved fruit quality through increasing flesh TSS percent, and contents of V. C. and sugars.

## CONCLUSION

In conclusion this study demonstrated that spraying the bio stimulant EM 30% had a similar effect to that of spraying with GA<sub>3</sub> at rate 30 ppm and a beneficial effect on growth and fruit set which had an impact on yield and fruit quality. Such approach is highly recommended

in confirming the need for safe food and sound environment in the light of an increasing demand for organic food.

### REFERENCES

- Abd-Ella, Eman, E.K. and Wafaa, A. A. Z. El-Sisi (2006).**Effect of foliar application of gibberellic acid and micronutrients on leaf mineral content, fruit set yield and fruit quality of Sultani fig trees. J. Agric. Res. Fac. Agric. Saba Basha, 11 (3):567-578.
- Addicott,F. T. and Addicott, A.B. (1982).** Abscission. Un,GA. Press. Lts. London, England, P.30-135.
- Association of Official Analytical Chemists A.O.A.C. (1995). Official methods of analysis, 15<sup>th</sup> ed. AOAC International Arlington, Verginia, USA.
- Atlas, R.M. and Bartha R. (1981).** Microbial Ecology: Fundamental and Applications. Addison-Wesley Publishing Company.560p.
- Bhanti,M. and Taneja, A. (2007).** Contamination of vegetables of different seasons with organophosphorous pesticides and related health risk assessment in northern India. Chemosphere 69: 63-68.
- Callejas-R; Bangerth, F.; Gardiola, J.L.; Garcia, J.L. and Quinlan J.D.(1998).** Is auxin export of apple fruit an alternative signal for inhibition of flower bud induction? Acta Hort. 463: 271-277.
- Chamberlain, T.P. and Daly, M.J. (2005).**Innovative use and adaption of microbial technology (EM) for large scale vegetable, arable and stock production on an organic farm in Canterbury, New Zealand. IFOAM conference Adelaide Australia 18-21.Septemper.
- Chapman, H.D. and Pratt, P.F. (1961).** Methods of Analysis for Soils, Plant and Water. Div. of Agric. Sci. Univ. Calif. pp: 309. USA.
- Desoky, I.M.; Riad, M. and Belatus, E. L. (2001).** Influence of effective microorganisms on growth and fruit characteristics of Papaya in Egypt. In Proceedings of the 6<sup>th</sup> International Conference on Kyusei Nature Farming. Ed. J. F. Parr et al., USDA, Washington DC.,USA.

- Donald, E.R.; Kathryn E.K.; Tahar, A. and Nicholas P.H.(2001).**Gibberellin regulates plant growth and development. A molecular genetic analyses of gibberellin signaling. Annual Review of Plant Physiology and Plant Molecular Biology, Vol.52:67-88.
- Dubois, M.; Gilles, K. A.; Homilton, J.K.; Robers, P.A. and Smith ,F. (1956).**Colorimetric methods for determination of sugar and related substances. Anal. Chem.28(3):350-458.
- El-Sabagh, A.S. and Mostafa, E.A.(2003).** Effect of gibberellic acid (GA<sub>3</sub>) treatments on vegetative growth, flowering density and fruiting of Anna apple cultivar. Alex. J. Agric. Res. 48 (2):75-86.
- El-Sabagh, A. S. and Ahmed,H.S.(2004).** Effect of gibberellic acid (GA<sub>3</sub>) and Sitofix on Anna apple crop load and fruit quality. Alex. J. Agric. Res. 49 (1): 71-79.
- Erkan, Z. and Bangerth,F.(1980).**Investigation on the effect of phytohormones and growth regulators on the transpiration, stomata aperture and photosynthesis of pepper (*Capsicum annum*) and tomato (*Lycopersicon esculatum Mull*). Plants Botany 54:207-220.
- Evenhuis, B. (1976).**Nitrogen determination Dept. Agric. Res. Royal. Tropical Inst. Amsterdam.
- Evenhuis, B. and DeWaard P.W.(1980)**Principles and practices in plant analysis. F.A.O. Soil Bull.39 (1): 152-162.
- Ferguson, J.J.; Avigne, W.T.; Allen, L.H. and Kocch, K.E. (1987).** Growth of CO<sub>2</sub> enriched sour orange seedlings treated with gibberellic and cytokinins. Proc. Florida State, Hort. Soc., 99:37-39.
- Grochowska, M.J.; Holdun, M.; Mika, A.; Morgas, H. and Chlebowska, D.(1995).** High responsiveness of apple trees to single application of growth regulator to the root collar. J. of Fruit and Ornamental Plant Research. 3: 91-100.
- Higa, T. (1995).**Effective microorganisms: Their role in Kyusei Nature Farming and Sustainable Agriculture In J.F.Parr ,S.B.Hornick, and C.E.Whitman (ed.) Proceedings of the First International

Conference of Kyusei Nature Farming U.S. Department of Agriculture, Washington, D.C.,USA.

- Higa, T. and Parr J. F.(1994).** Beneficial and effective microorganisms for a sustainable agriculture and environment. International Nature Farming Research Center, Atami, Japan,16 p -Higa,T. and Wididana, G. N.(1999). The concept and theories of microorganisms. In Proceedings of the First International Conference of Kyusei Nature Farming U.S. Department of Agriculture, Washington, D.C.,USA.
- Iwaishi, S. (2000).** Effect of organic fertilizer and effective microorganisms on growth, yield and quality of paddy-rice varieties. *J. Crop Prod* 3:269-273.
- Javaid, A. (2010).** Beneficial microorganisms for sustainable agricultural- A Review. In *Genetic Engineering, Biofertilisation, Soil Quality and organic Farming* Springer Publishers. DOI pp. 347-369.
- Javaid, A. and Shah, M. (2010).** Growth and yield response of wheat to EM (effective microorganisms) and parthenium green manure. *African J. of Biotechnology* Vol.9 (23), pp.3373-3381.
- Joo, Y.H.; Senanayake, Y.D. and Sangakkara, U.R.(1999).** Effect of EM on the production of crops and waste treatment in Korea . Fifth International Conference on Kyusei Nature Farming, Bangkok, Thailand, 23-26 October, 151-156 (c.f.CAB Abst.,1998).
- Kannaiyan, S. (2002).** Biofertilizers for sustainable production. In Kannaiyan, ed *Biotechnology of biofertilizers*. Narosa Publishing House, New Delhi. India, pp9-49.
- Legocka, J. (1987).** Kinetin induced changes in the population of translatable messenger RNA lacking a polyactenylated segment in cucumber cotyledons. *Acta Physiol;Plant* 9(1):33-39.(C.F.Plant Growth Reg. Abst., 13:1294).
- Malaka, S.M. (2008).** Effect of foliar application of liquid organic fertilizer (Aminofert), some micro-nutrients and gibberellin on

- leaf mineral content, fruit set, yield and fruit quality of Leconte pear trees. *Alex. J. Agric. Res.* 53 (1) 63-71.
- Murphy, J. and Riley, J.P.(1962).** A modified single solution method for the determination of phosphorus in natural water, *Anal Chem. Acta* 27:13-36.
- Nagodawithana, W.T. (1991).**Yeast Technology. Universal Food Corporation Milwaukee, Wisconsin. Published by Van Nostrand Reinhold, New York, p.273.
- Pati, B.R. and Chandra A.K. (1981).** Effect of spraying nitrogen-fixing phyllosphere bacterial isolates on wheat plants. *Plant and Soil* 61: 419-427.
- Piotr, Z. and Zofia Z. (2008).** Impact of biological effective microorganisms (EM) preparations on some physico-chemical properties of soil and the vegetative growth of apple trees rootstocks. *Nauka Przyroda Technologie. Dział Ogrodnictwo.* Pp 1-8.
- Ruinen, J.(1970).** The phyllosphere. The grass sheet, a habitat for nitrogen fixing microorganisms. *Plant and Soil* 33: 661-671.
- Sangakkara, U. and Marambe, B. (1998 ).** Influence of method of application of effective microorganisms on growth and yield of selected crops. In *Proceedings of the 4<sup>th</sup> International Conference on Kyusei Nature Farming.* Ed.J.F. Parr et al., USDA, Washington, DC, USA:111-117.
- Sangakkara, U. and Nissanka, S.P.(1998).** Impact of foliar application of effective micro-organisms on French beans. In *Proceedings of the 4<sup>th</sup> International Conference on Kyusei Nature Farming.* Ed.J.F. Parr et al., USDA, Washington, DC, USA: 321pp.
- Shaxson, T.F.(2006).** Re-thinking the conservation of carbon, water and soil: a different perspective. *Agron. Sustain. Dev.* 26: 9-19.
- Shou-Song, Y.; Cui-Ping, W.; Hui-Lian, X. and Jun-Ying, D.( 2002).** Effect of foliar application with effective microorganisms on leaf metabolism and seed yield in Soybean. *International Nature Farming Research Center, 5632 Hata Nagano, Japan.*

- Snedecor, G.W. and Cochran, W.G.(1990).** Statistical methods 7<sup>th</sup> ed. The Iowa. State. Univ. Press. Ames. Iowa,USA.p.593.
- Wididana, G. N. and Higa, T. (1998).** Effect of EM on the production of vegetable crops in Indonesia. In: Proceeding of the 4<sup>th</sup> International Conference on Kyusei Nature Farming, Bangkok,Thailand, 23-26.
- Wiltank, W. J. and Krezdorn, A. H. (1969).** Determination of gibberellins in ovaries and young fruits of navel oranges and their correlation with fruit growth. J. Amer. Soc. Hort. Sci.94:195-201.
- Wood, M. T.; Miles R. and Tabora, P. (1997).** EM fermented plant extract and EM5 for controlling pickleworm (*Diaphania nitidalis*) in organic cucumber. School of Natural Resources, University of Missouri, USA and EAREH College, Limon, Costa Rica.
- Xiaohou, S.; Diyou L.; Liang,Z.; Hu, W. and Hui, W.(2001).** Use of EM-technology in agriculture and environment management in China. Nat Farm Environ 2:9-18.
- Xu Hui-Lian.** (2000). Nature farming :History, principles and perspectives. J. of Crop Production.3 (1),1-10
- Yamada, K.and Xu HL (2000).** Properties and applications of an organic fertilizer inoculated with effective microorganisms. J. Crop Prod 3:255-268.
- Yousaf, Z.; Jilani G.Qureshi R.A. and Awan A.G. (2000).** Effect of EM on groundnut (*Arachis hypogaea L*) growth. Pak J. Biol.Sci.3:1803-1804.



## الملخص العربي

### تأثير الرش ببعض منشطات النمو علي نمو وإثمار وجودة ثمار أشجار التين السلطاني

ايمان السيد كامل عبد اللاه<sup>1</sup> - وفاء علي السيسى<sup>2</sup>  
 1- محطة بحوث البساتين بالصباحية - إسكندرية  
 2- الحديقة النباتية بالمعمورة - إسكندرية

أجري هذا البحث عامي 2009-2010 علي أشجار تين صنف سلطاني عمرها 10 أعوام ونامية في أرض جيرية بمنطقة برج العرب (حوالي 70 كم غرب الإسكندرية) لدراسة تأثير الرش بمنشط النمو الحيوي (EM) بتركيزات 10،20،30 % وكذلك منظم النمو حامض الجبريليك ( $GA_3$ ) بتركيزات 10،30،20 جزء في المليون علي النمو الخضري ونسبة عقد الثمار والمحصول وصفات جودة الثمار.

أوضحت النتائج عامة في كلا موسمي الدراسة أن جميع المعاملات لكل من المنشط الحيوي EM و  $GA_3$  ادت الى زيادة معنوية في النمو الخضري معبرا عنه بعدد النموات الحديثة وطول وقطر هذه النموات ومساحة الورقة وايضا نسبة العقد والمحصول (عدد، وزن الثمار/ شجرة) ومواصفات جودة الثمار (وزن وطول وقطر الثمرة، المواد الصلبة الذاتية، والسكريات الكلية) مقارنة بالأشجار الغير معاملة. ولم يكن للمعاملات أي تأثير علي محتوى الثمار من فيتامين ج .

واوضحت النتائج ان التركيز الاعلى لكل من EM و  $GA_3$  أعطى أفضل النتائج لجميع مواصفات النمو الخضري وأيضا نسبة العقد والمحصول.

وتحت ظروف هذه الدراسة يمكن التوصية باستخدام EM رشا علي اشجار التين بتركيز 30% لانتاج اعلى محصول مع افضل مواصفات جودة للثمار و كبديل امن عن استخدام منظم النمو حمض الجبريليك  $GA_3$ .



